

Vitreous: the resplendent enigma

J Sebag

Br J Ophthalmol 2009 93: 989-991 doi: 10.1136/bjo.2009.157313

Updated information and services can be found at: http://bjo.bmj.com/content/93/8/989.full.html

These include:

References	This article cites 24 articles, 6 of which can be accessed free at: http://bjo.bmj.com/content/93/8/989.full.html#ref-list-1
Email alerting service	Receive free email alerts when new articles cit this article. sign up in the box at the top right corner of the online article.
Topic collections	Articles on similar topics can be found in the following collections Retina (977 articles) Vitreous (102 articles)

Notes

To order reprints of this article go to: http://bjo.bmj.com/cgi/reprintform

Vitreous: the resplendent enigma

J Sebag

Arguably the most enigmatic of ocular structures, vitreous has long been unappreciated with respect to its role in health and disease. The classic anatomists and histologists of the 18th (Demours, 1741; Zinn, 1755) and 19th (Sir William Bowman, 1848; Virchow, 1885) centuries considered vitreous a tissue. However, in 1930, Duke-Elder¹ pointed out that vitreous is not a tissue but a cell product. This is consistent with our current concept of vitreous as an extended extracellular matrix composed primarily of water (98%), collagen and hyaluronan. The exact cells and processes responsible for the synthesis of these macromolecules are, as yet, unidentified. Also lacking is a comprehensive understanding of the supramolecular organisation of vitreous, limiting the ability of investigators and clinicians to study vitreous experimentally and evaluate it clinically. Indeed, even the roles of vitreous in ocular physiology are not universally established. One probable function of vitreous, however, is the maintenance of transparency within the eye (fig 1). This minimises light scattering and allows the unhindered transmission of photons to the retina for photoreception. Towards this end, vitreous possesses biochemical properties that inhibit cell migration and proliferation.^{2 3} While essential for vision, the invisibility of vitreous has obliged generations on both sides of the slit lamp and ophthalmoscope to merely look through vitreous, unable to adequately look at it.

The absence of effective diagnostic techniques with which to evaluate vitreous, both scientifically and clinically, has also hindered our ability to elucidate the role of vitreous in retinal pathology. For example, in the "Argument" of his 1930 BJO monograph on vitreous, ¹ Duke-Elder emphasised that "Inasmuch as the vitreous body is not a tissue but a cell product, its reaction to metabolic or toxic disturbances will be passive."

In particular, he proposed that it is passive gel liquefaction (and ensuing imbalances in "turgegescence" and "deturgescence") that detach the retina, even in the presence of a retinal break. Duke-Elder and his contemporaries failed to realise the paramount role of posterior vitreous detachment, a phenomenon that is not only acute, but unlikely passive. Recent advances in our knowledge of the molecular morphology of gel vitreous,^{4 5} the ultrastructure of the vitreoretinal interface⁶ and ageing changes in both^{7 8} have improved our understanding of the role of posterior vitreous detachment in retinal disorders.

Posterior vitreous detachment (PVD) is the most common acute event to occur in the human eye during the course of life. PVD is characterised by gel liquefaction (synchisis) and vitreoretinal dehiscence with collapse (syneresis) of the posterior vitreous away from the retina. PVD is considered by most to be an abnormal event. It is plausible, however, that much like the phenomenon of apoptosis, PVD is a beneficent part of Nature's "plan." It is known, for example, that in diseases such as diabetic retinopathy9 and age-related macular degeneration,¹⁰ complete PVD protects against more advanced stages of disease. Is it conceivable, therefore, that Nature has developed a way to mitigate the risk of advancing disease by salubriously separating vitreous away from



Figure 1 Human vitreous. The sclera, choroid and retina were dissected off the vitreous body, which remains attached to the anterior segment. Although placed on a surgical towel and exposed to room air, the shape of the vitreous body is spherical, and the gel is intact. The exquisite transparency of the vitreous body is evident. Specimen courtesy of the New England Eye Bank.

retina. Indeed, since PVD occurs without untoward sequelae in the overwhelming majority of individuals, it would appear that PVD is providential. In a minority of cases, however, liquefaction occurs without sufficient dehiscence at the vitreoretinal interface to allow for clean separation of the posterior vitreous cortex from the retina. This is known as "Anomalous PVD.""1 The sequelae of Anomalous PVD (fig 2) vary depending upon where the gel is most liquefied and where the posterior vitreous cortex is most firmly adherent to the retina. Also relevant is whether the posterior vitreous cortex that remains adherent to the retina is of full thickness or partial thickness. The latter is a consequence of a splitting in the posterior vitreous cortex, known as vitreoschisis.¹² Recent studies have further suggested that it may also matter whether the split occurs anterior or posterior to the level of the hyalocytes, the resident mononuclear phagocytes of the posterior vitreous cortex.13

The advent of new non-invasive imaging of the vitreoretinal interface has provided useful information regarding the fine structure of the posterior vitreous cortex in humans. As demonstrated in fig 3, there is a lamellar organisation of the collagen fibrils in the posterior vitreous of humans, confirming studies in monkeys.^{13 14} The lamellar structure within the posterior vitreous cortex constitutes a series of potential cleavage planes, making splits during anomalous PVD possible. Histopathological studies¹⁵ Moorfield's have shown that at vitreoschisis is present in 80% of eyes with proliferative diabetic retinopathy. Clinical investigations¹³ have detected vitreoschisis in half of eyes with macular holes and macular pucker. When vitreoschisis occurs. the outermost layer(s) of the posterior vitreous cortex remain(s) attached to the retina. Indeed, studies¹⁶ have shown that splitting of the posterior vitreous cortex can occur during vitreous surgery in 80% of eyes with macular pucker. It is likely that this phenomenon is at least partly if not wholly responsible for the recurrences observed following surgery for macular pucker.17

In this issue, Kifuku and colleagues *(see page 1016)* present their experience with 46 eyes undergoing surgery for so-called idiopathic epiretinal (this author prefers the term "premacular") membranes and no other confounding conditions like diabetic retinopathy, uveitis or trauma.¹⁸ Preoperatively, nearly all (96.7%) had

Correspondence to: Dr J Sebag, VMR Institute, 7677 Center Avenue, Suite 400 Huntington Beach, University of Southern California, Los Angeles, CA 92647, USA; jsebag@vmrinstitute.com



Figure 2 Anomalous posterior vitreous detachment (PVD). Schematic diagram of the various possible consequences of anomalous PVD. EXUD AMD, exudative age-related macular degeneration; VMTS, vitreomacular traction syndrome.

PVD, consistent with the findings of previous studies^{19–23} and with the concepts of anomalous PVD¹¹ and vitreoschisis.¹² After vitrectomy and membrane removal with intraocular forceps, the investigators stained the posterior pole with Brilliant Blue G to identify remnant tissue. Based on positive staining, they claimed to find "residual ILM" in 23/46 (50%) eyes. It is important to note that the uptake of this

dye is not definitive evidence that the remnant tissue was indeed the internal limiting lamina of the retina. Ten specimens were examined by flat-mount immunohistochemistry, which demonstrated that many cells, both immunopositive and immunonegative for glial fibrillar acid protein (GFAP), remained on this residual tissue. The true nature of this tissue cannot be ascertained by this



Figure 3 Lamellar structure of human posterior vitreous cortex. Three-dimensional optical coherence tomography imaging of the human postrior vitreous using the Spectralis (Heidelberg Engineering) with cinema 4DXL postprocessing. The left side of the image clearly demonstrates at least three layers in the posterior vitreous cortex. Courtesy of C Glittenberg and S Binder, Vienna, Austria.

histological approach, since a flat mount technique was employed with stains designed cells. for not tissue. Furthermore, if this tissue were indeed the ILM, why were so many cells present? These considerations all suggest, therefore, that this tissue is, at least in part, more likely the outer wall of a vitreoschisis cavity, with the GFAP-negative cells most likely representing hyalocytes and the GFAP positive cells of astrocytic origin.

The nature of the cells involved in macular pucker is controversial. Studies¹⁹⁻²³ have shown that nearly all eyes with macular pucker have a PVD; hence, it was believed that this caused breaks in the retina through which cells migrated and proliferated.23 However, histological examinations in a very large postmortem study failed to show breaks in the retina of any macular pucker eyes.²⁴ The concept of anomalous PVD with vitreoschisis proposes that during anomalous separation of vitreous from retina, there is a split within the posterior vitreous cortex anterior to the hyalocytes, leaving these cells embedded in the outer layer of the vitreoschisis cavity that is still attached to the macula.^{11–13} As was first hypothesised in 1989,25 hyalocytes have the capacity to stimulate cell migration, proliferation and membrane contraction, and could thus cause macular pucker. Scientific investigations have now provided evidence in support of this postulate.

Kohno and collaborators (see page 1020) studied 10 surgically excised premacular membranes from humans with macular pucker, and in separate experiments explored the effects of Transforming Growth Factor (TGF)-beta2 stimulation in an established in vitro wound-healing assay to compare the contractile properties of hyalocytes with those of human astrocytes.²⁶ Human vitreous removed at surgery for macular pucker was also tested for its ability to stimulate hyalocytes to induce gel contraction. The results showed that the predominant cells in the contracted portion of premacular membranes from macular pucker eyes are most likely hyalocytes. In vitro, hyalocytes were shown to induce considerable gel contraction, while astrocytes had no such effects. Vitreous samples from human eyes with macular pucker also stimulated gel contraction, an effect that was inhibited by anti-TGF-beta2-neutralising antibody. This and the other findings suggest that hyalocytes are pivotal in macular pucker pathogenesis and that TGF-beta 2 plays an important role in modulating the contractile phenomena. The authors suggest that further research will likely lead to pharmacotherapy that will modulate the behaviour of these critical cells. An alternative approach would be to induce total PVD prophylactically via pharmacologic vitreolysis,^{27–29} before the onset of anomalous PVD and its untoward sequelae.

All these recent advances belie the fact that not long ago, vitreous was a much maligned structure. The homogeneity of vitreous and lack of structures such as blood vessels and cells led anatomists to regard vitreous as "despicable," physiologists to consider it "negligible," and clinicians to see vitreous as "annoying."³⁰ Our new understanding of vitreous and our ability to manage its contribution to blindness will surely lead us to appreciate the resplendency of this enigma, for as put by Redslob,

Qu'y a-t-il de plus lumineux qu'un corps vitré

- Fraîchement décortiqué et exposé au soleil?
- [What could be more resplendent than a vitreus corpum
- Freshly excised and exposed to the sun?] E Redslob, Strasbourg, le 5 Janvier, 1932

Competing interests: None.

Br J Ophthalmol 2009;**93**:989–991. doi:10.1136/bjo.2009.157313

REFERENCES

- Duke-Elder WS. The nature of the vitreous body. Br J Ophthalmol 1930; (Monograph Suppl IV).
- Clamp AR, Jayson GC. The clinical potential of antiangiogenic fragments of extracellular matrix proteins. Br J Cancer 2005;93:967–72.
- Slevin M, Krupinski J, Gaffney J, et al. Hyaluronanmediated angiogenesis in vascular disease: uncovering RHAMM and CD44 receptor signaling pathways. *Matrix Biol* 2007;26:58–68.
- Sebag J. Macromolecular structure of vitreous. Prog Polym Sci 1998;23:415–46.
- Bishop PN. Structural macromolecules and supramolecular organisation of the vitreous gel. Prog Ret Eye Res 2000;19:323–44.
- Green WR, Sebag J. Vitreous and the vitreo-retinal interface. In: Ryan SJ, ed, *Retina*. St Louis: Mosby, 2001:Vol III, 1882–960.
- Sebag J. Age-related differences in the human vitreoretinal interface. Arch Ophthalmol 1991;109:966–71.
- Sebag J, Yee KMP. Vitreous—from biochemistry to clinical relevance. In: Tasman W, Jaeger EA, eds, *Duane's foundations of clinical ophthalmology*. Philadelphia, Lippincott Williams & Wilkins, 2007:Vol 1, Chapter 16.
- Akiba J, Arzabe CW, Trempe CL. Posterior vitreous detachment and neovascularization in diabetic retinopathy. *Ophthalmology* 1990;97:889–91.
- Krebs I, Brannath W, Glittenberg K, et al. Posterior vitreo-macular adhesion: a potential risk factor for exudative age-related macular degeneration. Am J Ophthalmol 2007;144:741–6.
- Sebag J. Anomalous posterior vitreous detachment: a unifying concept in vitreo-retinal disease. *Graefes Arch Clin Exp Ophthalmol* 2004;242:690–8.
- 12. Sebag J. Vitreoschisis. Graefes Arch Clin Exp Ophthalmol 2008;246:329–32.
- Sebag J, Gupta P, Rosen R, et al. Macular holes and macular pucker: The role of vitreoschisis as imaged by optical coherence tomography-scanning laser ophthalmoscopy. Trans Am Ophthalmol Soc 2007;105:121–31.
- Russell S, Hageman GS. Optic disc, foveal, and extrafoveal damage due to surgical separation of the vitreous. Arch Ophthalmol 2001;119:1653–8.

- Schwartz SD, Alexander R, Hiscott P, et al. Recognition of vitreoschisis in proliferative diabetic retinopathy. *Ophthalmology* 1996;103:323–8.
- Yamashita T, Uemura A, Sakamoto T. Intraoperative characteristics of theposterior vitreous cortex in patients with epiretinal membrane. *Graefes Arch Clin Exp Ophthalmol* 2008;246:333–7.
- Grewing R, Mester U. Results of surgery for epiretinal membranes and their recurrences. Br J Ophthalmol 1996;80:323–6.
- Kifuku K, Hata Y, Kohno R-i, *et al.* Residual internal limiting membrane in epiretinal membrane surgery. *Br J Ophthalmol* 2009;93:1016–19.
- Wise GM. Clinical features of idiopathic preretinal macular fibrosis. Am J Ophthalmol 1975;79:349–7.
- Sidd RJ, Fine SL, Owens SL, et al. Idiopathic preretinal gliosis. Am J Ophthalmol 1982; 94:44–8.
- Wiznia RA. Posterior vitreous detachment and idiopathic preretinal macular gliosis. *Am J Ophthalmol* 1986;102:196–8.
- Wang MY, Nguyen D, Hindoyan N, et al. Vitreopapillary adhesion in macular hole and macular pucker. Am J Ophthalmol. In press.
- Foos RY. Vitreoretinal juncture; epiretinal membranes and vitreous. *Invest Ophthalmol Vis Sci* 1977;16:416–22.
- Roth AM, Foos RY. Surface wrinkling retinopathy in eyes enucleated at autopsy. *Trans Am Acad Ophthalmol Otolaryngol* 1971;**75**:1047–58.
- Sebag J. The vitreous—structure, function, and pathobiology. New York: Springer, 1989:110–11.
- Kohno R-i, Hata Y, Kawahara S, et al. Possible contribution of hyalocytes to idiopathic epiretinal membrane formation and its contraction. Br J Ophthalmol 2009;93:1020–6.
- Sebag J. Pharmacologic vitreolysis. *Retina* 1998;18:1–3.
- Sebag J. Is pharmacologic vitreolysis brewing? Retina 2002;22:1–3.
- Sebag J. Molecular biology of pharmacologic vitreolysis. Trans Am Ophthalmol Soc 2005;103:473–94.
- Redslob E. Le Corps Vitré—son développement, sa structure, ses propriétés physico-chimiques. Rapport de la Societé Française d'Ophtalmologie, Masson et Cie, Paris, 1932:V (preface).